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Blood 142 (2023) 6819-6820

The 65th ASH Annual Meeting Abstracts

## **ONLINE PUBLICATION ONLY**

## 703.CELLULAR IMMUNOTHERAPIES: BASIC AND TRANSLATIONAL

## Generation of Hinge-Modified Human CD19 Chimeric Antigen Receptor (CAR) T Cells for the Treatment of CD19-Low Leukemia

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CAR T cell therapy targeting CD19 has emerged to show spectacular success in the treatment of B leukemias and lymphomas. This strategy is mainly achieved by potentiating T-cells to recognize and target tumor cells. However, CD19 CAR T cell therapy can result in neurotoxicity<sup>1</sup> and cytokine-associated toxicity as well as relapse with antigen dim or negative disease <sup>2</sup>; therefore, strategies to mitigate these side effects are needed.

Recent trials in adults with lymphoma using fully human CAR (Hu19-28) T cells resulted in the secretion of lower levels of cytokines and conferred lower neurologic toxicity. <sup>3</sup> To assess the potential responsiveness of this Hu19-28 CAR T cells against B-acute lymphocytic leukemia (B-ALL), T cells engineered with the Hu19-28 CAR were compared with murine FMC63-based CARs. Hu19-28 and FMC63-based CAR T cells exhibited a rapid increase in IFNg secretion in response to the CD19 <sup>+</sup> NALM6 ALL cell line (CD19 <sup>High</sup>). Interestingly, and consistent with the lower toxicity of Hu19-28 CAR T cells in patients with lymphoma, IFNg secretion by Hu19-28 CAR T cells in response to a CD19 <sup>Low</sup> NALM6 line was significantly reduced as compared to FMC63-28 CAR T cells. Hu19-28 CAR T cells also exhibited high *in vitro* cytotoxicity against CD19 <sup>High</sup> NALM6. Notably though, Hu19-28 CAR T cells demonstrated lower *in vitro* cytotoxicity against CD19 <sup>Low</sup> NALM6 as compared to FMC63-28 CAR T cells. These important differences were also detected *in vivo*; both Hu19-28 and FMC63-28 CAR T cells eradicated CD19 <sup>High</sup> leukemia in an NSG mouse model but only the latter efficiently controlled the *in vivo* growth of CD19 <sup>Med</sup> leukemia.

As we and others have shown that the hinge domain of the CAR plays a critical role in its function, especially against low antigen density tumors <sup>4,5,6</sup>, we evaluated the impact of the hinge on the function of the Hu19-28 CAR T cells. Compared with the initial construct with a CD8a hinge length of 54 amino acids (Hu19-8H <sup>54</sup>-28), Hu19 constructs with 45 (Hu19-H <sup>45</sup>-28), 25 (Hu19-H <sup>25</sup>-28), and 13 (Hu19-8H <sup>13</sup>-28) amino acids were generated. All CAR constructs were highly expressed at the surface of transduced T lymphocytes and were efficient in the killing of CD19 <sup>High</sup> leukemia. However, only the Hu19-H <sup>25</sup>-28 and Hu19-8H <sup>13</sup>-28 CAR constructs exhibited high cytotoxicity against CD19 <sup>Low</sup> leukemia (**Figure 1**). Together these data support the preclinical evaluation of Hu19-28 CAR constructs with shorter CD8a hinge lengths for the treatment of pediatric patients with relapsed/refractory ALL.

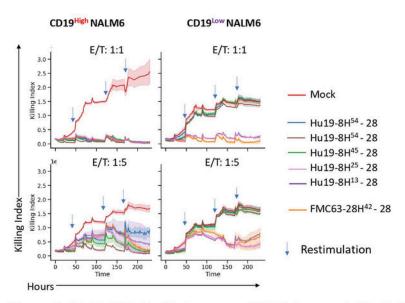
References

1. Gust J et al (2018) CNS Drugs, 32(12) 1091-1101. https://doi.org/10.1007/s40263-018-0582-9

2. Lee DW et al (2019) Biology of Blood and Marrow Transplantation, 25(4) 625-638. https://doi.org/10.1016/j.bbmt.2018.12.758 3. Brudno JN et al (2020). Nature Medicine, 26(2) 270-280. https://doi.org/10.1038/s41591-019-0737-3

4. Majzner RG et al (2020) Cancer Discovery, 10(5), 702-723. https://doi.org/10.1158/2159-8290.CD-19-0945

Chen X/ Mirazee JM *et al* (2022) Journal of Magnetic Resonance 340, 107234. https://doi.org/10.1016/j.jmr.2022.107234 Mirazee et al., in preparation **Disclosures Kochenderfer:** Kite, a Gilead company: Research Funding; Bristol Myers Squibb: Research Funding. **Shah:** Lentigen: Research Funding; CARGO: Consultancy; VOR: Consultancy, Research Funding; Immunoadoptive Cell Therapy Private Limited: Consultancy, Other: Scientific Advisory Board.



**Figure 1.** <u>Hu19-CAR T with truncated CD8 hinges exhibit high *ex vivo* cytotoxicity against CD19<sup>Low</sup> leukemia. Healthy donor T lymphocytes were transduced with HuCD19 CARs with different size CD8 $\alpha$  hinges as well as the canonical FMC63 murine CAR with a CD28 $\alpha$  hinge. The indicated CART and untransduced T cells (mock) were co-cultured with GFP+ NALM6 lines harboring high (CD19<sup>High</sup>, left) or low (CD19<sup>Low</sup>, right) levels of CD19 at either a 1:1 or 1:5 effector/target (E/T) ratios and repeated NALM6 stimulations every 48h-72h as depicted by the arrows. *Ex-vivo* cytotoxicity was monitored using the Incucyte<sup>TM</sup> system over 200 hours and killing index over time is presented.</u>

Figure 1

https://doi.org/10.1182/blood-2023-187954